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# Two new triterpenoids from the resin of Boswellia carterii

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#### Two new triterpenoids from the resin of Boswellia carterii

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Two new triterpenoids, 3-oxotirucalla-7,9(11),24-trien-21-oic acid (1) and  $18H\alpha$ ,3 $\beta$ ,20 $\beta$ -ursanediol (2), along with 15 known triterpenes,  $\alpha$ -amyrin,  $\alpha$ -boswellic acid,  $\beta$ -boswellic acid, acetyl  $\alpha$ -boswellic acid, acetyl  $\beta$ -boswellic acid, 9,11-dehydro- $\beta$ -boswellic acid, 9,11-dehydro- $\alpha$ -boswellic acid, acetyl 11 $\alpha$ -methoxy- $\beta$ -boswellic acid, 11-keto- $\beta$ -boswellic acid, acetyl 11-keto- $\beta$ -boswellic acid, acetyl  $\alpha$ -elemolic acid, 3 $\beta$ -hydroxytirucalla-8,24-dien-21-oic acid, elemonic acid, 3 $\alpha$ -hydroxytirucalla-7,24-dien-21-oic acid, and 3 $\alpha$ -hydroxytirucall-24-en-21-oic acid, were isolated from the resin of *Boswellia carterii* Birdw.

**Keywords:** triterpenoids; *Boswellia carterii* Birdw (Burseraceae); 3-oxotirucalla-7,9(11),24-trien-21-oic acid;  $18H\alpha$ ,  $3\beta$ ,  $20\beta$ -ursanediol

#### 1. Introduction

Boswellia carterii Birdw is an arbor distributed in Somalia and Ethiopia. Its resin was used to treat coronary heart disease, angina pectoris, and ulcer in China [1]. Previous research for the resin showed the presence of a number of triterpenes including oleanane-, ursane-, and tirucallane-type [2,3]. Boswellic acidtype triterpenes have been reported to inhibit the activity of 5-lipoxygenase and topoisomerases [4-6] and to induce apoptosis in several types of tumor cells including colon, prostate, and malignant glioma [7-10]. As our continuing work on searching bioactive triterpenes from natural medicines, chemical investigation of the resin of B. carterii led to the isolation of two new triterpenes (1 and 2) and 15 known triterpenes (Figure 1). The known triterpenes were identified as  $\alpha$ -amyrin [11],  $\alpha$ -boswellic acid [2],  $\beta$ -boswellic acid [2], acetyl  $\alpha$ -boswellic acid [2], acetyl  $\beta$ -boswellic acid [2], 9,11-dehydro- $\beta$ boswellic acid [3], 9,11-dehydro- $\alpha$ -boswellic acid, acetyl 11a-methoxy-B-boswellic acid [12], 11-keto- $\beta$ -boswellic acid [13], acetyl 11-keto- $\beta$ -boswellic acid [2], acetyl  $\alpha$ -elemolic acid [13], 3 $\beta$ -hydroxytirucalla-8,24-dien-21-oic acid [13], elemonic acid [13],  $3\alpha$ -hydroxy-tirucalla-7,24-dien-21-oic acid [3], and  $3\alpha$ hydroxytirucall-24-en-21-oic acid [3] by comparison of their spectroscopic data with those of published values. Details of the isolation, structure elucidation of compounds 1 and 2, and their antileukemia activities are presented here.

#### 2. Results and discussion

Compound 1 was obtained as colorless needles, and its molecular formula was

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Figure 1. The structures of compounds 1-4.

determined as  $C_{30}H_{44}O_3$  on the basis of its HREIMS spectrum (m/z 451.32041) $[M - H]^+$ ). The <sup>1</sup>H NMR spectrum showed seven methyl singlets and the <sup>13</sup>C NMR spectrum revealed 30 carbon signals, which were sorted by DEPT experiment as seven methyls, eight methylenes, six methines, and nine quaternary carbons, of which six  $sp^2$  carbons and two carbonyl groups were suggested based on the chemical shifts (see Table 1). Due to tirucallane-type as only a tetracyclic triterpenoid isolated from the resin in previous studies, compound 1 was presumed as a tirucallane-type triterpene in view of biogenetic source. The <sup>13</sup>C NMR spectrum indicated the presence of two carbonyl groups, resonating at  $\delta$  214.9 (C-3) and 178.5 (C-21), two conjugated double bonds at  $\delta$  119.0 (C-7), 141.6 (C-8), 144.2 (C-9), and 116.6 (C-11), and a single double bond at  $\delta$  124.8 (C-24) and 131.8 (C-25). The gross structure of compound 1 was deduced from its HMBC spectrum. The conjugated double bond positions at C-7(8) and C-9(11) were determined by the long-range correlations between the methyl group at  $\delta$  1.07 and carbons at  $\delta$  50.4 (C-5) and 144.2 (C-9) and between the methyl group at  $\delta$  0.96 and carbons at δ 141.6 (C-8), 49.7 (C-14), 31.2 (C-15), and 44.5 (C-13). The HMBC correlations of H-26 ( $\delta$  1.64) and H-27 ( $\delta$ 1.60) with C-24 ( $\delta$  124.8) and C-25 ( $\delta$ 131.8) indicated a double bond located at C-24. The tirucallane-type triterpene had the same tetracyclic skeleton as a lanostane-type triterpene, but the orientation of the methyl group at C-13 and the side chain at C-17 was different from that in lanostane-type triterpene, i.e. the tirucallane-type triterpene had the  $\alpha$ -orientation for both the methyl group and the side chain, while the lanostane-type had  $\beta$ orientation for both of them. The <sup>13</sup>C NMR spectral data of compound 1 are extremely analog with (20§)-3-oxolanostain 7,9(11),24-trien-21-oic acid (dehydrotrametenonic acid, 3) [14], which easily lead to mistake compound 1 as the above lanostane-type one (3). The relative stereochemistry was determined based on the NOESY experiment. The NOESY correlations of H-30/H-19, H-19/H-28, H-

	1			2	
	$ \begin{aligned} \delta_{\rm H} \left( J,{\rm Hz} \right) \\ \left( 600{\rm MHz} \right) \end{aligned} $	$\delta_{C}$ (75 MHz)	$\begin{array}{c} \text{HMBC} \\ (\text{H} \rightarrow \text{C}) \end{array}$	$\frac{\delta_{\rm H} \left( J,{\rm Hz} \right)}{\left( 600{\rm MHz} \right)}$	δ <sub>C</sub> (75 MHz)
1	1.91 m, 1.64 m	37.0	C-19	1.68 m	38.6
2	2.76 td (12.5, 5.2), 2.29 m	35.0	C-1, 3	1.64 m, 1.57 m	28.5
3		214.9		3.24 dd (10.9, 5.2)	79.0
4		47.8			38.8
5	1.67 dd (11.4, 4.8)	50.4	C-4, 10, 19, 29	0.69 m	55.0
6	2.10 m, 1.42 m	27.3	C-5, 7, 8, 10	1.51 m, 1.37 m	18.3
7	5.40 br s	119.0	C-14	1.40 m	34.3
8		141.6			41.2
9		144.2		1.20 m	49.4
10		36.4			36.9
11	5.14 br s	116.6	C-10	1.46 m, 1.52 m	21.3
12	2.42 m	36.4	C-9, 11, 13, 14	1.66 m, 1.58 m	27.4
13		44.5		1.82 td (12.0, 4.3)	38.8
14		49.7			43.0
15	1.74 m, 1.40 m	31.2	C-8, 13, 14, 30	1.33 m, 1.21 m	26.5
16	2.10 m, 2.06 m	24.4		1.34 m, 1.17 m	38.3
17	2.48 q (10.8)	48.2	C-20		35.6
18	1.00 s	16.6	C-12, 13, 14	1.05 m	47.9
19	1.07 s	19.9	C-5, 9, 10	1.53 m	41.9
20	2.62 td (11.1, 3.2)	48.8			75.0
21		178.5		1.51 m	37.8
22	1.91 m, 1.75 m	33.2		1.27 m, 1.21 m	40.2
23	2.36 m, 2.26 m	26.7	C-24, 25	0.97 s	28.0
24	5.31 t (7.0)	124.8	C-26, 27	0.77 s	15.4
25		131.8		0.84 s	16.1
26	1.64 s	25.8	C-24, 25, 27	1.04 s	16.0
27	1.60 s	17.7	C-24, 25, 26	0.94 s	14.6
28	0.96 s	24.8	C-8, 13, 14, 15	0.90 s	18.3
29	1.07 s	22.1	C-3, 4, 5, 30	1.06 d (6.0)	17.4
30	1.03 s	23.4	C-3, 4, 29	1.09 s	21.4

Table 1. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectral data of compounds **1** and **2** in CDCl<sub>3</sub>.

28/H-17, and H-18/H-20 indicated the H-17 on the  $\beta$ -face and the methyl group at C-13 on the  $\alpha$ -face of the molecule, which confirmed the side chain at C-17 in  $\alpha$ orientation and the tirucallane-type skeleton of compound **1** (Figure 2). So, compound **1** was characterized as 3-oxotirucalla-7,9(11),24-trien-21-oic acid.

Compound **2** was obtained as colorless needles, and its molecular formula was determined as  $C_{30}H_{52}O_2$  on the basis of its HREIMS spectrum at m/z 444.3961 [M]<sup>+</sup>.



Figure 2. Key NOESY correlations of compounds 1 and 2.

The <sup>1</sup>H NMR spectrum exhibited eight methyl signals at  $\delta$  1.09 (3H, s, H-29), 1.06 (3H, d, J = 6.0 Hz, H-30), 1.04 (3H, s, H-26), 0.97 (3H, s, H-23), 0.94 (3H, s, H-27), 0.90 (3H, s, H-28), 0.84 (3H, s, H-25), 0.77 (3H, s, H-24), and an oxygenated methine signal at  $\delta$  3.24 (H-3, dd, J = 10.9, 5.2 Hz). The <sup>13</sup>C NMR spectrum showed 30 carbon atoms, which were sorted by DEPT experiment as eight methyls, ten methylenes, six methines, and six quaternary carbons. An oxygenated quaternary carbon resonating at  $\delta$  75.0 and an oxymethine carbon at  $\delta$  79.0 were exhibited, and no olefinic carbon was observed in the <sup>13</sup>C NMR spectrum. Based on the above data, compound 2 was deduced as an ursane-type triterpene. The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra are similar to the corresponding data of 3B,20Sdihydroxytaraxastane (4) [15] except the chemical shift values at C-19, 29, 30, and suggested that the C-19 in compound 2 may be of R-configuration, which was confirmed by the analysis of coupling constants and the NOESY experiment. H-13( $\beta$ ) signal resonating at  $\delta$  1.82 (1H, td,  $J = 12.0, 4.3 \,\mathrm{Hz}$ ) together with the NOESY correlation between H-18 and H-27 revealed the axial  $\alpha$ -orientation of H-18. The NOESY correlations for H-25 with H-26, H-26 with H-13, H-13 with H-28, and H-28 with H-29 indicated that they are on the  $\beta$ -face. So, compound 2 was assigned as  $18H\alpha$ ,  $3\beta$ ,  $20\beta$ -ursanediol. The <sup>1</sup>H and <sup>13</sup>C NMR spectral data were assigned by the <sup>1</sup>H-<sup>1</sup>H COSY, HSQC, and HMBC spectra (Table 1).

#### 3. Experimental

#### 3.1 General experimental procedures

Melting points (uncorrected) were measured on a Yanaco MP-S3 micromelting point apparatus. Optical rotations were measured with a Perkin-Elmer 241MC polarimeter. IR was conducted on a Perkin IFS-55 spectrometer. NMR spectra were recorded on a Bruker ARX 300 NMR spectrometer and a Bruker ARX 600 NMR spectrometer. The chemical shifts were quoted relative to TMS, and the coupling constants were in Hz. DEPT, HMBC, HSQC, COSY, and NOESY were measured on a Bruker ARX 600 NMR spectrometer. EIMS (70 eV) was conducted on a Shimadzu GCMS-QP5050A spectrometer. HREIMS were recorded on an Autospec-UltimaETOF instrument. The chromatographic silica gel (200–300 mesh) was produced by Qingdao Ocean Chemical Factory (Qingdao, China), and Sephadex LH-20 was bought from GE Healthcare (London, England).

#### 3.2 Plant material

The resin of *B. carterii* was bought from Liaoning Medicinal Material Corporation, Shenyang, China and identified by Prof. Qishi Sun of Shenyang Pharmaceutical University. A voucher specimen (20070918) has been deposited in the Department of Natural Products Chemistry, Shenyang Pharmaceutical University, Shenyang, China.

#### 3.3 Extraction and isolation

The resin of B. carterii (1000 g) was extracted with CHCl<sub>3</sub>. After removing the solvent, the extract (678 g) was chromatographed on a column of silica gel with gradient elution using petroleum ether with increasing proportions of EtOAc to give four fractions (Parts 1-4). Part 3 (150 g) was separated by column chromatography on silica gel, eluted with EtOAc/petroleum ether (8:2-7:3) to produce 59 fractions. Subfractions 8-19 were combined and separated by silica gel H column chromatography and eluted with acetone/petroleum ether (100:10, 100-15) to give compound 2 (4 mg). Part 4 (87 g) was separated by column chromatography on silica gel, eluted with EtOAc/petroleum ether (7:1-2:1) to produce 113 fractions. Subfractions 1-35 were combined and chromatographed on a column of Sephadex LH-20 eluted with MeOH followed by open ODS column to give compound **1** (6 mg).

#### *3.3.1 3-Oxotirucalla-7,9(11),24-trien-21-oic acid (1)*

Colorless needles (acetone), mp 267–269°C,  $[\alpha]_{20}^{20}$  – 40.0 (c = 0.4, CHCl<sub>3</sub>). IR (KBr)  $\nu_{max}$  3440 (OH), 1703 (COOH), 1695 (CO), 1652 (C=C) cm<sup>-1</sup>. <sup>1</sup>H and <sup>13</sup>C NMR spectral data (see Table 1). HREIMS: m/z 451.3204 [M – H]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>43</sub>O<sub>3</sub>, 451.3212).

#### 3.3.2 $18H\alpha$ , $3\beta$ , $20\beta$ -Ursanediol (2)

Colorless needles (acetone), mp 293–295°C,  $[\alpha]_D^{20} - 33.0$  (c = 0.3, CHCl<sub>3</sub>). IR (KBr)  $\nu_{\text{max}}$  3558 (OH), 2921 and 2855 (CH), 1463 (CH) cm<sup>-1</sup>. <sup>1</sup>H and <sup>13</sup>C NMR spectral data (see Table 1). EIMS: m/z (rel. int) 444 [M]<sup>+</sup> (2.7), 374 (52), 207 (35), 189 (51). HREIMS: m/z 444.3961 [M]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>52</sub>O<sub>2</sub>, 444.3967).

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#### References

 H. Guo and L. Zhang, *Food Drug* 9, 50 (2007).

- [2] J.Y. Zhou and Y. Cui, *Acta Pharm. Sin.* 37, 633 (2002).
- [3] N. Banno, T. Akihisa, K. Yasukawa, H. Tokuda, K. Tabata, Y. Nakamura, R. Nishimura, Y. Kimura, and T. Suzuki, *J. Ethnopharmacol.* 107, 249 (2006).
- [4] H. Safayhi, T. Mack, J. Sabieraj, M.I. Anazodo, L.R. Subramanian, and H.P. Ammon, J. Pharmacol. Exp. Ther. 261, 1143 (2002).
- [5] D. Poecke and O. Werz, Curr. Med. Chem. 13, 3359 (2006).
- [6] R.F. Hoernlein, T. Orlikowsky, C. Zehrer, D. Niethammer, E.R. Sailer, T. Simmet, G.E. Dannecker, and H.P.T. Ammon, *J. Pharmacol. Exp. Ther.* 288, 613 (1999).
- [7] J.J. Liu, B.H. Huang, and S.C. Hooi, *Br. J. Pharmacol.* 148, 1099 (2006).
- [8] T. Syrovets, J.E. Gschwend, B. Büchele, Y. Laumonnier, W. Zugmaier, F. Genz, and T. Simmet, *J. Biol. Chem.* 280, 6170 (2005).
- [9] M. Lu, L.J. Xia, H.M. Hua, and Y.K. Jing, *Cancer Res.* 68, 1180 (2008).
- [10] Y.S. Park, J.H. Lee, J.A. Harwalkar, J. Bondar, H. Safayhi, and M. Golubic, *Adv. Exp. Med. Biol.* 507, 387 (2002).
- [11] S.B. Mahato and A.P. Kundu, *Phytochem-istry* 37, 1517 (1991).
- [12] S. Schweizer, A.F.W. von Brocke, S.E. Boden, E. Bayer, H.P.T. Ammon, and H. Safayhi, J. Nat. Prod. 63, 1058 (2000).
- [13] R.S. Pardhy and S.C. Bhattacharyya, *Indian J. Chem.* **16B**, 174 (1978).
- [14] T. Akihisa, Y. Mizushina, and M. Ukiya, *Biosci. Biotechnol. Biochem.* 68, 448 (2004).
- [15] G.S. Susunaga, A.C. Siani, M.G. Pizzolatti, R.A. Yunes, and F. Delle Monache, *Fitoterapia* 72, 709 (2001).